# EFFECT OF LINSEED OIL BEADS ADDITION WITH VITAMIN E ON PERFORMANCE, BLOOD METABOLITES AND MILK YIELD OF LACTATING GOATS

Abeer M. El-Essawy<sup>1</sup>, I.M. Khattab<sup>2</sup>, Ahlam R. Abdou<sup>1</sup> and A.M. Abdel-Wahed<sup>1</sup>

<sup>1</sup>Animal and Poultry Nutrition Department, Desert Research Center, El-Matarya, Cairo, Egypt.

<sup>2</sup> Departments of Animal and Fish Production, Faculty of Desert and Environmental Agriculture, Matrouh University, 51744 Matrouh, Egypt.

Correspondence Author: Abeer M. El-Essawy Animal and Poultry Nutrition Department, Desert Research Center, El-Matarya, Cairo, Egypt. E-mail: - abeerateek@hotmail.com

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# SUMMARY

his study aimed to determine the effects of addition of capsulated linseed oil (beads) alone or in combination with vitamin E ( $\alpha$ -tocophyrol acetate) as feed additives on animal performance, blood plasma and milk fatty acid (FA) profiles during lactation period. Twenty four Damascus pregnant doses of about 44.5 ± 1.2 kg live body weight were randomly allotted to 3 groups of 8 doses each and assigned to receive one of three experimental diets: control group: where goats were received berseem hay and concentrate feed mixture (50:50) without any additives, (CON), LO group: goats were offered control diet plus 2.5 gm beads/head/day and LO+VE group: goats were offered control diet plus 2.5 gm beads plus 600 IU VE/head/day. The experiment lasted for about 135 days (from mid pregnancy and lasted 60 days post-partum). Results showed that LO beads inclusion strongly increased milk yield, fat (P<0.01), protein, lactose, total solids yield (P<0.05) and milk fat % (P<0.05) compared with control. Blood plasma  $\gamma$ -linolenic acid was positively affected by beads while C20:2 $\omega$ 6 was increased (P<0.05) by beads plus VE. Moreover, beads inclusion reduced lauric (C12:0), myristic (C14:0) and palmitic (C16:0) (P<0.05) acids while increased contents of monounsaturated fatty acids (MUFA) as oleic (C18:1 $\omega$ 9) and polyunsaturated fatty acids (PUFA) as  $\gamma$ -linoleic (C18:3 $\omega$ 3) (P<0.01) and C18:4 $\omega$ 3 (PP<0.05) compared with control milk. As a consequence reduction of the proportions of total SFA (P<0.01), total omega-6 FAs (P<0.05) and omega-6: omega-3 ratio and enhanced the proportions of unsaturated FAs (UFA) and omega-3 FAs (P<0.01) resulted in health quality of goat's milk was improved. In addition, some of blood plasma metabolites were improved with beads where plasma createnine, cholesterol (PP<0.05) and lipase activity (P<0.01) were decreased compared with control animals. Plasma urea (P<0.05), triglycerides (TG) (P<0.01), low density lipoprotein (LDL) (P<0.05), high density lipoprotein (HDL) (P<0.01) and total antioxidant capacity (TAC) (P<0.05) were increased while lipase activity was decreased with VE addition. The inclusion of VE combined with LO beads didn't result in benefits in goat's products. Further in vivo studies should be undertaken to explore suitable level of vitamin E in association with protected form of useful and healthy fatty acids.

Keywords: Linseed, fatty acid profile, milk, vitamin E and goats.

# **INTRODUCTION**

Animal products provide a considerable percentage of saturated fats that affecting human health. Nutrition constitutes a natural way to modulate animal's products fatty acids (FA) composition. Addition of linseeds to ruminant diets is useful to increase the concentration of polyunsaturated FA in dairy products and meat. Ruminant products contain a variety of FA, particularly polyunsaturated fatty acids (PUFA), which are considered beneficial to human health (Doreau and Ferlay, 2015). The inclusion of linseed as a source of PUFA in the diet has been used to enhance these beneficial FA in animal products (Abuelfatah *et al.*, 2016). Milk fatty acid profile is influenced by dietary fatty acids and ruminal fatty acid

metabolism including lipolysis and biohydrogenation of UFA (Bai *et al.*, 2018). Thus, the use of fat sources protected from rumen biohydrogenation could be a more practical approach (Scott *et al.*, 1971).

Many attempts aimed to protect PUFA against rumen biohydrogenation to improve dairy products and meat fatty acid content. So, different protection methods have been applied such as encapsulation of lipids in a coat of proteins treated with formaldehyde (Doreau et al., 2011; and Fievez et al., 2007). However, this technique is not used, because of it's high cost, and possible adverse effects of excessive amounts of polyunsaturated fatty acids on animal health and product quality (Doreau and Ferlay, 2015). Commercially supplementation with FA calcium salts is used although saturated and monounsaturated (MUFA) react with calcium forming an insoluble product that resist rumen biohydrogenation but PUFA do not react to form calcium salts then no protection against biohydrogenation of PUFA (Lundy et al., 2004; and Castañeda-Gutiérrez et al., 2007). Therefore, Gawad et al. (2015) found a new encapsulation method using biopolymers to protect FAs in linseed oil from rumen biohydrogenation. An increase in milk PUFA content increased it's susceptibility to oxidation, so it may require dietary supply of antioxidants to prevent oxidative damage of the fatty acids. Vitamin E being one of the most powerful antioxidants, it plays an important role in the prevention of lipid oxidation in cell membranes (Deaville et al., 2004). Vitamin E is used in animal feed mainly due to its lipid-soluble antioxidant function and because it assists in reducing the effects of oxidative stress. Therefore, the uses of dietary antioxidants with dairy animals increase the oxidative stability of milk (Santos et al., 2014).

The overall objective of the current study is to determine the effects of protected lipid supply alone or in combination with vitamin E on Damascus does performance during lactation period and on blood plasma and milk fatty acid profiles.

## MATERIALS AND METHODS

#### Preparing linseed oil beads:

Encapsulated linseed oil was prepared according to the method described by Gawad *et al.* (2015). Briefly, 2.5% (w/v) alginate/k-carrageenan solution was prepared by dissolving sodium alginate/k-carrageenan gel (1:1 w: w) in distilled water using an overhead mechanical stirrer. Then, linseed oil emulsion gel was prepared by mixing linseed oil with alginate/carrageenan solution (20%; v/v) using Tween 80 as an emulsifier (0.5 ml /100 ml gel). Uniform linseed oil beads were prepared using Encapsulator instrument (model IE-50 R was purchased from Encap. Biosys., Switzerland). Linseed emulsion gel solution was injected in encapsulater and let solution for dripping under specific conditions as follow: Nozzle: 1ml, frequency 1700 HZ, flow rate: 4 ml min<sup>-1</sup> and air pressure of 1 bar. The formed beads were received in 2.5% CaCl<sub>2</sub> (w/v) and left in hardening solution for up to 30 min for more hardening. Then, linseed oil beads were filtered and dried using oven dryer (45 °c).

#### Fatty acids profile of linseed oil and linseed oil beads:

Fatty acids profile of linseed oil before and after encapsulation was analyzed according to AOAC (2000) using Ultra Gas Chromatographs.

#### Animals, diets and experimental design:

Twenty-four Damascus goats in mid pregnant stage with an initial body weight (BW) of  $44.5 \pm 1.2$  kg were divided into three groups. Goats within groups (8 goats each) were assigned randomly to one of three treatments. The three dietary treatments were; control treatment (CON) where the animals were fed basal diet without any additive. The second; linseed oil beads treatment (LO) where the experimental animals were fed basal diet plus a daily dose of 2.5 g /goat/d of linseed oil beads. The third, LO+VE, the animals were fed basal diet plus a daily dose of 2.5 g /goat/d of linseed oil beads + 600 IU /goat/d of vitamin E ( $\alpha$ -tocopherol acetate). The daily dose of vitamin E was orally administered once daily directly after feeding with the aid of a syringe. The experimental animals had free access to water and mineral block. Goats with their kids were housed in individual concrete pens, fed individually, and milked. The experiment was extended from mid pregnancy and lasted 60 days post-partum. The experimental diets were offered once daily at 08.00 h. The beads were top dressed and mixed with a fork into the basal diet. Before starting the experiment, goats were vaccinated against internal and external parasites and intro-toxemia. Live body weights of does and their kids were measured biweekly and body weight changes were calculated.

#### Feed intake and chemical analysis:

Feed intake was measured throughout the experimental period by weighing the offered diets and refusals from the previous day. The feed samples were dried in a forced-air oven at 65 °C for 72 h, and ground in a Willey mill with a 1.0-mm screen. Samples were analyzed for DM (method 930.15), ash (method 942.05), Nitrogen (method 954.01) and ether extract (EE; method 920.39), according to AOAC, (1997) official methods. Neutral detergent fiber was determined by the procedure of Van Soest *et al.* (1991) without use of an alpha amylase but with sodium sulfite and expressed exclusive of residual ash. Acid detergent fiber was analyzed according to AOAC, (1997) and expressed exclusive of residual ash. The chemical composition of the basal diet is shown in Table (1).

Item	Concentrate feed mixture	Berseem hay		
Dry matter	90.45	88.32		
Organic matter	97.16	87.65		
Crude protein	17.32	13.22		
Ether extract	4.25	2.96		
Ash	2.82	12.35		
Neutral detergent fiber	17.82	55.38		
Acid detergent fiber	10.80	38.01		

Table (1): Nutrient composition of the basal diet (% on DM basis).

### Rumen liquor sampling and analysis:

At the end of the experiment, ruminal contents were sampled randomly from four goats in each group at 4 h after the morning feeding by stomach tube. Approximately 100 mL of rumen fluid were collected and strained through 4 layers of cheesecloth. The pH of ruminal fluid was measured immediately with a pH meter (Accumet Model 15, Fisher Scientific, USA). A 50-mL aliquot of ruminal fluid was acidified with 2.5 mL of 6 N HCl and frozen ( $-20^{\circ}$ C) for subsequent determination of ammonia-N concentration calorimetrically using commercial kit (Konitzer and Voigt, 1963) and total volatile fatty acids (TVFA) (Warner, 1964).

### Milk sampling, milk composition and fatty acid analysis:

Milk production was determined biweekly as follow: kids were separated from their dams, then does were milked completely by hand at 8.00 and 18.00 h and kids were return to their dams from the second week of lactation till the end of the experiment. The collected milk was recorded, and a subsample was taken from individual does at each milking and mixed as a constant percentage of the morning and eve ning to obtain the sample of each does, then stored at -20 °C until further analysis. Milk samples were analyzed for total solids, fat, protein, and lactose using infrared spectrophotometry (Foss 120 Milko-Scan, Foss Electric, Hillerød, Denmark). Yield of fat-corrected milk was calculated according to the equation reported by Gravert (1987): FCM (3.5%) = 0.433MY + 16.218 FY, where: FCM: fat-corrected milk; MY: milk yield (kg/day); FY: fat yield (kg/day).

#### Blood plasma sampling and analysis:

At the end of the experiment, 10-mL blood samples were taken before morning feeding from the jugular vein of each animal into a clean tube containing EDTA. The plasma were separated by centrifuging at 3000 rpm for 20 minutes and frozen at -20 °C up to subsequent analysis. Blood plasma samples were analyzed using commercial kits. All plasma samples were analyzed for total protein, albumin, globulin (by subtraction the total proteins values from the albumin values), urea –N, createnine, total lipids (TL), cholesterol (TC), triglycerides (TG), high density lipoprotein (HDL), low density lipoprotein (LDL), lipase enzyme, total antioxidant capacity (TAC), alanine amino transferase (ALT) and aspartate amino transferase (AST) using Biodiagnostic laboratory kits.

#### Milk and plasma fatty acids analysis:

The milk samples were pooled per does, resulting in one milk sample per does during the experiment. Milk lipids were determined according to the method of AOAC (2000), where FAs are methylated with boron trifluoride in methanol, extracted with heptane and determined on a gas chromatograph with FID detector (PE Auto System XL) with auto sampler and Ezchrom integration system. Oven temperature 200 °C, injector and detector 250 °C. On the other hand, plasma lipids were extracted by using 100% ether /

sample in the ratio of 1:10 (v/v) as detailed by Ferraz *et al.* (2004) then methylated and determined by the same procedure referred to milk FAs.

#### Statistical analysis:

The DMI, milk yield, fat-corrected milk and milk composition data were analyzed as repeated measurements over time (weeks) using mixed procedure of SAS (2006) (Statistical Analysis System, version 9.2 for Windows; SAS Institute, Cary, NC, USA). The model used was the following: Yij=  $\mu$  + Di + bj + Hk + Di × Hk + eij; where, Yij=dependent variable;  $\mu$  =overall mean; Di=effect of diet i, i=1 to 3; bj=effect of block j, j=1 to 8; Hk=effect to the week k, k=2 to 8; Di×Hk=interaction of diet i×week k; eij=random effect. The initial BW, final BW, BW change of the does and kids throughout the experiment, rumen parameters, blood plasma parameters and the FA profile of the milk fat and of plasma were analyzed using MIXED procedure of SAS (Statistical Analysis System, version 9.2 for Windows; SAS Institute, Cary, NC, USA). The model used was the following: Yij =  $\mu$  + Di + bj + eij, where  $\mu$  = overall mean, Di = effect of diet, bi = effect of block and eij = random effect. Orthogonal contrasts were used to compare the effects of: linseed oil beads (CON vs. LO) and the association between linseed oil beads and vitamin E (LO vs. LO+VE). The effects were considered significant when P < 0.05.

# **RESULTS AND DISCUSSION**

#### Fatty acids composition of beads:

Results of Table (2) indicated that encapsulation process did not affect FAs content of oil especially

Table (2): Fatty acids composition of linseed oil and linseed oil beads.	Table (	2)	: Fatty	v acids com	position	of lin	iseed oil	and	linseed	oil beads.
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Fatty acids %	Linseed oil	Linseed oil beads
SFA		
C6:0, Caproic	0.63	0.67
C8:0, Caprlyic	0.33	0.37
C14:0, Myristic	0.25	0.27
C15:0, Pentaenoic	0.18	0.2
C16:0, Palmitic	7.46	7.98
C17:0, Heptadecanoic	0.19	0.14
C18:0, Stearic	5	3.9
C20:0, Arachidic	0.33	0.24
C22:0, Behenic	0.3	0.36
∑Saturated fatty acids MUFA	14.67	14.13
C15:1\u00fc6, 10-Pentadecanoic	0.48	0.48
C16:1ω7, Palmitoleic	0.1	0.21
C18:1ω9, Oleic	18.43	18.41
C18:1007, Vaccinic	0.94	0.97
C20:1ω9, Gadolic	0.3	0.27
C20:1ω7, 9-eicosaenoic	ND	0.15
C20:1w5, 11-eicosaenoic	ND	0.25
C22:1009, Erucic	0.21	0.45
∑Monounsaturated PUFA	20.46	21.19
C18:2006, Linoleic	13.47	14.1
С18:2ω4,	ND	0.18
C18:3ω3, Linolenic	51.4	50.4
$\sum$ Poly unsaturated fatty acids	64.87	64.68
Unsaturated fatty acids	85.33	85.87
Omega-3 FA	51.4	50.4
Omega-6 FA	13.95	14.58

ND: non detectable

PUFA fractions. Therefore, linolenic (C18:3 $\omega$ 3) and linoleic (C18:2 $\omega$ 6) acids which are the most important PUFA were not affected by encapsulation process. Their values were 51.4 and 50.4% for linolenic acid and 13.47 and 14.1% for linoleic acid in linseed oil and it's beads, respectively. The present results are in agreement with Gawad *et al.* (2015), who used the same technology of encapsulation.

Table (3): Effect of supplementation	of	linseed	oil	beads	with	or	without	vitamin	Е	on	does
performance.											

Itam		Treatment	1	SEM	P- value <sup>2</sup>			
Item –	CON	LO	LO + VE	SEM	CON vs. LO	LO vs. VE		
No. of does	8	8	8					
Body weight of does, kg								
Initial	44.06	44.56	44.81	1.20	0.901	0.949		
Final	41.71	42.38	42.75	1.23	0.872	0.927		
Body weight changes	-2.46	-2.19	-2.06	0.09	0.147	0.630		
Dry matter Intake, kg	1.73	1.71	1.69	0.07	0.864	0.833		
Production, g/d								
Milk	1574	1807	1824	35.68	0.001	0.794		
FCM <sup>3</sup>	1484	1802	1798	43.77	0.003	0.960		
Fat	49.49	62.85	62.15	1.78	0.005	0.810		
Protein	49.27	59.47	56.87	1.46	0.016	0.440		
Lactose	71.92	84.27	85.87	2.10	0.013	0.729		
Total solids	183.5	222.4	219.9	5.89	0.010	0.846		
Milk composition, %								
Fat	3.14	3.48	3.41	0.05	0.033	0.340		
Protein	3.15	3.29	3.12	0.04	0.194	0.153		
Lactose	4.56	4.67	4.71	0.07	0.645	0.742		
Total solids	11.64	12.30	12.06	0.15	0.213	0.420		

1Treatments: Control group fed on basal diet without additives; LO group: basal diet with linseed oil beads; LO+VE group: fed on basal diet with linseed oil beads and vitamin E. 2 CON vs. LO: control diet vs basal diet and linseed oil beads; LO vs. VE: basal diet + linseed oil beads vs basal diet + linseed oil beads + vitamin E. SEM, standard error of means.

 Table (4): Effect of supplementation of linseed oil beads with or without vitamin E on sulking kids.

Item	Treatme	ent <sup>1</sup>		— SEM	P- value <sup>2</sup>	P- value <sup>2</sup>				
Item	CON	LO	LO + VE		CON vs. LO	LO vs. VE				
No. of kids	11	11	11							
Birth weight, kg	3.80	3.84	3.77	0.15	0.925	0.760				
Body weight, kg										
After 30 days	8.54	8.71	8.79	0.17	0.443	0.766				
After 60 days	13.51	14.13	14.41	0.26	0.374	0.648				
Average daily gain, g/	Average daily gain, g/d									
After 30 days	158.1	162.6	167.3	6.13	0.697	0.659				
After 60 days	165.6	180.3	187.3	6.79	0.422	0.685				

<sup>1</sup>Treatments: Control group fed on basal diet without additives; LO group: basal diet with linseed oil beads; LO+VE group: fed on basal diet with linseed oil beads and vitamin E. <sup>2</sup> CON vs. LO : control diet vs basal diet and linseed oil beads; LO vs. VE : basal diet + linseed oil beads vs basal diet + linseed oil beads + vitamin E. SEM, standard error of means.

#### Milk production and composition:

Milk yield was increased with LO supplied goats compared with CON goats (Table 3) and this increment agree with findings of Gomez – Cortes *et al.* (2009), Benchaar *et al.* (2012) and Kholif *et al.* (2015), who used different forms of linseed or it's oil. This increase in milk production may be attributed to that LO resulted in higher volatile fatty acids concentration in the rumen of the supplemented goats as matched with the following present results in Table (4). Moreover, LO supply may affect mammary gland metabolism. Petit (2003) illustrated that greater milk production could be a result of greater dietary amino

acids available for absorption by the animal. Crawford and Hooverm (1984) and Petit (2003) agreed with the present data. On the other hand, Zened et al. (2012), Suksombat et al. (2014) and Almeida et al. (2019) found that milk yield did not affect by oil diet with or without VE, Petit et al. (2005) recorded a reduction in milk production. All of the macro components yield were affected by LO supply compared with milk of CON goats. Milk fat corrected milk (FCM) was increased by LO treatment comparing with milk of CON goats (225.1 vs 185.6), respectively. Milk fat yield and it's percentage were increased with LO supply and this may be attributed to effective oils protection against ruminal bio hydrogenation increased fat yield (Ashes et al., 1992) while ineffective protection (Petit et al., 2002) or low level of added fat (Tymchuk et al., 1998) had no effect on milk fat yield. Similarly, Knapp et al. (1991); Kim et al. (1993) and Petit and Gagnon, (2009) observed higher yield and percentage of milk fat when cows were fed whole oilseeds or protected fats. This may be due to increased dietary FA being taken up by mammary gland for milk fat synthesis (Knapp et al., 1991). Generally, supplemental fat may elevate milk yield and milk fat but sources and types of fat affecting differently on such parameters (Chouinard et al., 1997). Elevated concentrations of milk protein with LO supply may be due to increased oil supply of amino acids for synthesis of milk protein. Petit (2002) and Petit (2003) were in agreement with the present findings. Kholif et al. (2014) attributed the increase of milk protein to improvement of ruminal microbial protein synthesis. Some of the previous studies such as Chilliard et al. (2009) and Radivojević et al. (2011) reported no change in milk protein whereas, Pires et al. (1996) and Miller et al. (2009) found that milk protein percentage was decreased particularly for cows fed on extruded oilseeds. In the same line, the lactose production was increased with LO treatment. Similar result was reported by Petit (2003) with formaldehyde treated oilseeds while, Almeida et al. (2019) suggested that lactose production is rarely changed by diet. All of the significant effects noted previously with LO beads supply were suppressed when vitamin E supplied with beads. Focant et al. (1998) reported that vitamin E supply increased the concentration of this vitamin in milk by 45% resulting in higher resistance of such milk fat to oxidation.

### Kids growth rates:

No differences were identified between lactating doses fed on either LO or LO+VE (p>0.05) and control ones regarding their kids birth weight or change in body weight till weaning (Table 4). These observations in terms of average daily gain are consistent with the results of the previous studies on growing lambs supplemented with different vegetable oils (Miltko *et al.* 2019), fattening lambs fed on soybean meal or extruded linseed (Facciolongo *et al.* 2018) and weaned male lambs of Tan sheep supplemented with vitamin E (Zhao *et al.* 2013).

#### **Ruminal parameters:**

Results of Table (5) indicated comparable levels of ruminal ammonia (p>0.05) on addition of LO or LO + VE while feeding of linseed oil beads increased the concentration of volatile fatty acids (VFA's) (P<0.05) compared to control goats while their concentrations were decreased (P<0.01) when VE incorporated compared to LO fed goats. These results may be due to the anaerobic fermentation of linseed oil or crushed linseed that increased yielding of VFA's (Kholif *et al.*, 2015) and / or to improvement of ruminal fermentation due to enhancement of cellulolytic bacteria and protozoa activities with linseed (El–Essawy, 2019). In contrast, Kim *et al.* (2007) in sheep, Benchaar *et al.* (2012) in dairy cows and Abuelfatah *et al.* (2016) in goats demonstrated no effect of linseed or linseed oil supplementation on pH and VFA's. These differences in concentrations of TVFA were probably attributed to differences in rumen volume (Chikunya *et al.*, 2004) or to changes in the rumen species composition of microbes in response to inclusion of unsaturated fatty acids in linseed that being toxic to cellulolytic and methanogenic bacteria (Broudiscou *et al.*, 1994).

Table (5): Effect of supplementation	of	linseed	oil	beads	with	or	without	vitamin	E on	rumen
fermentation.										

Item	Treatm	ent <sup>1</sup>		SEM	P- value <sup>2</sup>			
Item	CON	LO	LO + VE	– SEM	CON vs. LO	LO vs. VE		
рН	6.11	6.22	6.21	0.02	0.050	0.891		
Ammonia-N, mg /dl	18.13	19.74	16.85	0.241	0.499	0.224		
Volatile fatty acid, mg /dl	7.1	8.14	6.49	1.42	0.018	0.0003		

<sup>1</sup>Control, basal diet without oil supplement; LO, basal diet with linseed oil beads; LO+VE basal diet with linseed oil beads and vitamin E. <sup>2</sup> CON vs. LO = control diet vs diet supplemented with linseed oil; LO vs. VE = diet supplemented with linseed oil vs diet supplemented with linseed oil and vitamin E. SEM, standard error of the means.

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An increased tendency of rumen pH is recorded (p=0.05) with LO addition compared to CON values whereas VE inclusion did not change pH values compared to those fed on LO. Numerically higher ruminal pH on feeding LO was in agreement with previous studies using different lipid sources including fish oil in diets of sheep (Wachira *et al.*, 2000), in diet of cows (Shingfield *et al.*, 2003) and both of these studies attributed this increase to associated decreases in DMI and also in diets of cattle fed cottonseed alone or combined with vitamin E (Nogueira *et al.*, 2019) and they explained higher pH to higher NDF content of cottonseed. On the other hand, Gawad *et al.* (2015) recorded lower ruminal pH values in vitro with different levels of linseed oil beads but they suggested that the mean of pH values remained within normal range of pH which is between (6 and 6.7). Other studies reported a lack of change in pH values with whole linseed inclusion compared to a protected palm oil (Scollan *et al.*, 2001) or compared to soybean (El–Essawy, 2019).

### Blood plasma metabolites and fatty acids:

Regarding LO supply in feeding resulted in decreasing blood plasma createnine, cholesterol (TC) (P<0.05) and lipase activity (P<0.01) (Table 6). The reduced cholesterolemia probably attributed to enrichment of linseed with omega-3 fatty acids. These results are in harmony with El-Essawy (2019) who found a reduction in createnine and TC with linseed supply due to enrichment of linseed with omega-3 fatty acids. In the same line, Weill *et al.* (2002) observed a repeated decrease in fat content of animal products as a result of inhibition of lipogenesis by alpha linolenic acid (Price *et al.*, 2000) and also linoleic acid that known to reduce TC (Weill *et al.*, 2002). Another study showed that oilseed supplementation increased the concentration of TC in bovine blood (Gonthier *et al.*, 2005) and these variations may be attributed to different sources and quantity of oilseed (Liu *et al.*, 2008). However, LO supply allowed lower lipase activity may be attributed to increase cellulolytic bacterial population hence both bacteria and fungi are predominant microbial sources of lipase (Sangeetha *et al.* 2011). Vitamin E supply, resulted in elevated levels of urea, triglycerides (TG), low – density lipoprotein (LDL), high –

measurements.						
	Treatment <sup>1</sup>				P- value <sup>2</sup>	
Item	CON	LO	LO +	SEM	CON vs. LO	LO vs. VE
			VE			
Total protein, g/dl	7.89	9.02	10.50	0.62	0.487	0.331
Albumin, g/dl	3.83	3.90	3.62	0.13	0.651	0.545
Globulin, g/dl	4.05	5.12	6.89	0.68	0.508	0.293
Urea, mg/dl	67.63	58.75	70.6	2.45	0.235	0.026
Creatinine, mg/dl	1.44	0.98	0.86	0.09	0.019	0.211
Total lipid, mg/dl	75.17	74.74	74.79	0.75	0.863	0.804
Triglycerides, mg/dl	10.50	11.36	20.43	0.25	0.549	0.003
Cholesterol, mg/dl	88.33	74.41	83.18	2.07	0.010	0.137
LDL-cholesterol, mg/l	40.69	39.60	43.73	0.55	0.224	0.019
HDL-cholesterol, mg/dl	63.42	63.73	72.90	1.39	0.878	0.009
Lipase, U/l	150.0	116.7	91.33	8.66	0.005	0.013
Total antioxidant capacity,	0.334	0.350	0.497	0.02	0.656	0.014
mM/l						
Aspartate amino	16.2	15.2	15.6	0.44	0.446	0.740
transferase, U/l						
Alanine amino transferase,	16	17.4	17.6	0.53	0.206	0.923
U/l						

 Table (6): Effect of supplementation of linseed oil beads with or without vitamin E on blood plasma measurements.

<sup>1</sup>Control, basal diet without oil supplement; LO, basal diet with linseed oil beads; LO+VE basal diet with linseed oil beads and vitamin E. <sup>2</sup> CON vs. LO = control diet vs diet supplemented with linseed oil; LO vs. VE = diet supplemented with linseed oil vs diet supplemented with linseed oil or of means. density lipoprotein (HDL) and total antioxidant capacity while lipase activity was considerably decreased.

El–Essawy (2019) supported our findings where she reported increased blood urea due to improved CP digestibility with linseed inclusion whereas Sharma *et al.* (1972) found reduced values of blood urea were related to decreased N degradability. Petit (2003) found that blood urea was not affected by different oilseeds treated with formaldehyde.

#### Fatty acid composition in blood plasma and milk:

# Saturated fatty acids (SFA):

The SFA in blood plasma were not affected significantly (p>0.05) by any of feed additives, (Table 7) although most of them showed insignificant decrease on feeding of LO compared to CON goats as capric (C10:0), undecanoic (C11:0), tridecanoic (C13:0), myristic (C14:0), palmitic (C16:0) and heptadecanoic (C17:0) acids. As a consequence of this reduction, the total SFA was decreased numerically in LO fed goats compared to CON ones (33.79 vs. 37.82) instead vitamin E supply, increased total SFA compared to LO fed group (36.39 vs. 33.79, respectively) indicating that BH of PUFA and its transformation to SFA was decreased with LO addition. So, many studies by Chikunya *et al.* (2004) and Fiorentini *et al.* (2015) concluded that the process of bio hydrogenation is the main cause of reduction of PUFA leaving the rumen. Consequently, SFA's especially lauric (C12:0), myristic (C14:0) and palmitic (C16:0) acids were also decreased (P<0.05) therefore, total SFA's were decreased (P<0.01) significantly in goat's milk fed LO treatment (Table 8). Chichlowski *et al.* (2005) and Liu *et al.* (2008) found similar trend with different oilseeds sources. Indeed, reduction of SFA in animal products especially C14:0 and C16:0 with LO supply prevent many physiological problems for consumer such as elevated blood cholesterol (Suksombat *et al.*, 2014) because C16:0 (palmitic acid) has hypercholesterolemic properties (Kennelly, 1996).

profile (g of fatty acid/ 100 g) in blood plasma.											
	Treatmer	nt <sup>1</sup>		_	P- value <sup>2</sup>						
Fotty poid %	CON	LO	LO +	SEM	CON	vs.	LO	vs.			
Fatty acid %	CON	LU	VE		LO		VE				
SFA											
C10:0, capric	0.51	0.22	0.22	0.18	0.423		0.994				
C11:0, undecanoic	2.8	2.3	2.08	0.23	0.478		0.837				
C12:0, lauric	0.94	1.22	0.8	0.22	0.776		0.253				
C13:0, tridecanoic	1.91	1.84	1.96	0.58	0.965		0.957				
C14:0, myristic	2.43	1.71	1.51	0.27	0.379		0.256				
C15:0, pentadecanoic	1.56	1.71	2	0.14	0.758		0.224				
C16:0, palmitic	8.01	7.41	10.91	0.84	0.658		0.289				
C17:0, heptadecanoic	12.09	9.7	9.09	0.76	0.275		0.855				
C18:0, stearic	7.57	7.68	7.83	0.24	0.897		0.778				
$\sum$ Saturated	37.82	33.79	36.39	1.04	0.414		0.375				
<u>–</u> MUFA											
C18:1w7, vaccinic	0.8	1.17	2.09	0.38	0.368		0.431				
C18:1ω9, oleic	7.91	7.7	8.88	0.82	0.881		0.757				
∑Monounsaturated	8.71	8.87	10.97	1.16	0.928		0.666				
PUFA											
C16:4ω3	13.96	15.36	12.97	0.8	0.442		0.394				
C18:2w6, linoleic	15.56	15.39	15.56	0.54	0.918		0.882				
C18:3ω3, ylinolenic	1.64	5.29	5.28	0.64	0.036		0.995				
C18:4ω3, alpha Octadecatetraenoic	10.03	9.94	10.14	0.51	0.942		0.903				
C20:2ω6	6.5	7	5.94	0.35	0.698		0.031				
C20:4\u03, ecosatartrienoic	4.18	3.87	1.85	0.8	0.891		0.182				
∑Polyunsaturated	52.66	56.85	51.74	1.66	0.48		0.089				
$\sum$ Unsaturated	61.37	65.72	62.71	1.27	0.484		0.5				
Other fatty acids	0.81	0.49	0.9	1.29	0.607		0.377				
∑Omega-6	22.06	22.39	21.5	0.31	0.652		0.399				
$\overline{\Sigma}$ Omega-3	30.6	34.45	30.87	1.81	0.521		0.237				
Omega-6: omega-3	0.79	0.65	0.7	0.06	0.52		0.55				

Table (7): Effect of supplementation of linseed oil beads with or without vitamin E on fatty acids profile (g of fatty acid/ 100 g) in blood plasma.

<sup>1</sup>Control, basal diet without oil supplement; LO, basal diet with linseed oil beads; LO+VE basal diet with linseed oil beads and vitamin E. <sup>2</sup> CON vs. LO = control diet vs diet supplemented with linseed oil; LO vs. VE = diet supplemented with linseed oil vs diet supplemented with linseed oil and vitamin E.SEM, standard error of the means.  $\sum$  Saturated fatty acids: total saturated fatty acids,  $\sum$  Unsaturated fatty acids: total unsaturated fatty acids,  $\sum$  Poly unsaturated fatty acids: total poly unsaturated fatty acids fatty acids.

#### Monounsaturated fatty acids (MUFA):

Vaccinic acid (C18:1 $\omega$ 7) was increased numerically (p=0.36) in blood plasma (Table 7) and in milk (p= 0.25) (Table 8) with beads supply compared to control goats (1.17 *vs.* 0.8) in blood and (0.30 *vs.* 0.22) in milk, respectively indicating an incomplete biohydrogenation by microbial activity (Corl *et al.*, 2001). Also, vaccinic acid is known as the main precursor of conjugate linoleic acid (CLA), which is an isomers group of linoleic acid that has the capacity to perform several metabolic benefits in human as inhibition of lipogenesis, blood cholesterol, cancer incidence reduction and changes in muscle tissues growth (Rice *et al.*, 2012). Almeida *et al.* (2019) found the same trend of the present results with fish oil supplemented to goats and also, oleic acid was increased (P<0.01) in milk in the same group. Oleic acid (C18:1 $\omega$ 9) concentration as a main MUFA was not significantly affected by dietary additives in blood whereas, it increased (P<0.01) in milk of goats fed LO compared to CON goats. This could be due to higher uptake from blood and/ or higher synthesis from stearic acid as observed by Bas *et al.* (2007). As a result, the concentration of total MUFA was not affected in blood (p>0.05) however, it increased (P<0.01) in milk of goats fed on LO compared to CON goats. These results are in agreement with Gawad *et al.* (2015) who used batch culture system and demonstrated that protected forms of linseed oil (beads) didn't affect MUFA levels after 24h of incubation.

profile (g of fatty acid/ 100 g of milk fat) in milk.										
		Treatme	nt <sup>1</sup>		_	P- value <sup>2</sup>				
Fatty acid %		CON	LO	LO + VE	SEM	CON vs. LO	LO vs. VE			
SFA										
C8:0, caprlyric		2.05	1.89	1.6	0.09	0.392	0.348			
C10:0, capric		7.73	6.25	6.21	0.29	0.083	0.757			
C12:0, lauric		3.74	3.1	2.91	0.13	0.049	0.421			
C14:0, myristic		8.87	8.18	8.34	0.11	0.015	0.495			
C15:0, pentadecanoic		1.58	1.44	1.38	0.05	0.213	0.661			
C16:0, palmitic		27.88	26.99	27.28	0.15	0.016	0.52			
C17:0, heptadecanoic		1.95	2.19	2.1	0.08	0.291	0.584			
C18:0, stearic		15.87	15.21	15.16	0.16	0.186	0.807			
C20:0, arachidic		0.23	0.24	0.28	0.01	0.391	0.108			
$\sum$ Saturated fatty acid		69.9	65.49	65.26	0.68	0.008	0.558			
MUFA										
C16:1ω7, palmitoleic		1.67	1.59	1.46	0.05	0.663	0.404			
C18:1007, vaccinic		0.22	0.3	0.34	0.02	0.257	0.416			
C18:1ω9, oleic		24.49	28.98	28.89	0.66	0.001	0.859			
∑Monounsaturated PUFA		26.38	30.87	30.69	0.66	0.001	0.738			
C16:3\u03c64, hexagonic		0.26	0.33	0.28	0.01	0.116	0.317			
C18:2\u00f36, linoleic		2.39	2.28	2.37	0.05	0.521	0.682			
C18:3 $\omega$ 3, y-linolenic		0.19	0.37	0.33	0.02	0.001	0.147			
- 0	alpha									
Octadecatetraenoic	uipiiu	0.23	0.37	0.38	0.02	0.028	0.892			
C20:4 $\omega$ 6, arachidonic		0.29	0.2	0.17	0.02	0.118	0.238			
$\Sigma$ Polyunsaturated		3.36	3.55	3.53	0.05	0.229	0.908			
$\sum$ Unsaturated fatty acid		29.74	34.42	34.22	0.68	0.001	0.689			
Other fatty acids		0.36	0.09	0.52	0.46	0.079	0.735			
$\sum$ Omega-6		2.68	2.48	2.53	0.04	0.048	0.569			
$\sum \text{Omega-3}$		0.42	0.74	0.71	0.05	0.004	0.559			
Omega-6: omega-3 ratio		6.45	3.37	3.59	0.44	0.005	0.582			

 Table (8): Effect of supplementation of linseed oil beads with or without vitamin E on fatty acids profile (g of fatty acid/ 100 g of milk fat) in milk.

IControl, basal diet without oil supplement; LO, basal diet with linseed oil beads; LO+VE basal diet with linseed oil beads and vitamin E. 2 CON vs. LO = control diet vs diet supplemented with linseed oil; LO vs. VE = diet supplemented with linseed oil vs diet supplemented with linseed oil oil; standard error of the means.  $\sum$  Saturated fatty acids: total saturated fatty acids,  $\sum$  Unsaturated fatty acids: total unsaturated fatty acids;  $\sum$  Poly unsaturated fatty acids: total poly unsaturated fatty acids.

#### Polyunsaturated fatty acids (PUFA):

Linseed oil beads treatment resulted in greater concentration of linolenic acid (C18:3ω3) in blood plasma (P<0.05) (Table 7) and in milk (P<0.01) (Table 8) compared to CON group. This finding was expected as a consequence of its high dietary level in oil beads composition (Table 2) where part of this FA passed through the rumen without suffering biohydrogenation, then incorporated into the milk. Results of Almeida et al. (2019) agree with the present finding. The linoleic acid (C18:206) supply for blood plasma was comparable among treatments (p>0.05), consequently, the concentration of this acid was not changed on goat's milk among treatments (Table 8). Although, alpha octadecatetraenoic (C18:4 $\omega$ 3) acid was not affected in blood plasma but it's concentration was increased (P<0.05) in milk on LO feeding compared to control group (0.37 vs. 0.23), respectively. Kholif *et al.* (2015) concluded that feeding dairy goats with fat source affecting indirectly lipogenesis in the mammary glands. The total PUFA proportion was numerically influenced by LO beads in blood plasma and in milk (p>0.05). There were no significant differences between dietary groups in blood plasma (p>0.05) regarding the total of omega-6, omega-3 or omega-6/omega-3 ratio while in milk, the total omega-6 and omega-6/omega-3 ratio were decreased significantly (P < 0.05 and P < 0.01, respectively) with a desirable increase in concentration of total omega-3 (P<0.01) due to beneficial effects of this FA for human health (Ruxton, 2007) compared to CON milk. Gawad et al. (2015) proved that the content of PUFA, omega-3 and omega-6 were increased with beads supply and decreased ratio of omega-6/ omega-3 after incubation with batch culture system. The higher concentrations of unsaturated fatty acids (UFA) supplied for the blood plasma (P>0.05) and milk (P>0.01) of goats supplied with LO may be attributed to greater fat content in feed additives resulting in increased absorption of UFA then transferred from diets to milk (Almeida et al., 2019). Thus, it is a considerable gain in nutritional value of milk in terms of UFA especially omega-3 fatty acids contents. A similar trend was recorded in the recent study of Almeida et al. (2019) with different oilseeds sources. Lastly, an effect of vitamin E on FA profiles is limited. Chikunya et al. (2004) consistent with the current results where they proved that vitamin E supplementation to the sheep diet didn't affect the efficiency of rumen BH nor the trans C18:1 proportion among FAs.

# CONCLUSION

Encapsulated beads supply as a source of polyunsaturated fatty acids did not affect the DMI, dose weight, birth weight and weight after 60 days of kids but increased milk production and the yield of all macro compounds of it as fat corrected milk, fat, protein, lactose and total solids and fat% only. Also, dietary supply of PUFA increased TVFAs, changed the FAs profile in blood plasma and in milk. Linseed oil beads promoted the increment of unsaturated FA particularly omega-3 FA and reduced SFA, omega-6 FA and omega6: omega-3 ratio in milk. Consequently, milk FA profile was improved and became healthier for consumers. However, the inclusion of vitamin E combined with linseed oil beads did not result in benefits to goat's milk. Further in vivo studies should be undertaken to explore suitable level of vitamin E in association with protected form of useful and healthy fatty acids to improve milk FA profile.

### REFERENCES

- Abuelfatah, K., A.B. Zuki, Y. M. Goh, A. Q. Sazili and A. Abubakr (2016). Effects of feeding whole linseed on ruminal fatty acid composition and microbial population in goats. J. Animal Nutrition., 2:323-328.
- Almeida, O.C., J. r. M. V. C. Ferraz, I. Susin, R.S. Gentil, D. M. Polizel, E. M. Ferreira, J.P.R. Barroso and A.V. Pires (2019). Plasma and milk fatty acid profiles in goats fed diets supplemented with oils from soybean, linseed or fish. J. Small Ruminant Research 170: 125–130.
- AOAC (1997). Official Methods of Analysis. 16th Ed. Assoc. Offic. Anal. Chem., Arlington, VA.
- AOAC (2000). Official Methods of Analysis. Association of Official Analytical Chemists. 17 Ed. 969.3 and 991.39 Fatty acids in oils and fats preparation of Methyl esters boron tri fluoride – AOAC-IUPAC method codex – Adopted – AOAC method. Chapter 41, 19-20.

### Egyptian J. Nutrition and Feeds (2019)

- Ashes J.R., B.D. Siebert, S.K. Gulati, A.Z. Cuthbertson and T.W. Scott (1992). Incorporation of n-3 fatty acids of fish oil into tissue and serum lipids of ruminants, Lipids 27: 629–631.
- Bai, S., Z. Cao, B. Cao, H. Yang, S. Li and J. Liu (2018). Effects of different forage combinations in total mixed rations on in vitro gas production kinetics, ruminal and milk fatty acid profiles of lactating cows. J. Anim Sci., 89:1261–1270.
- Bas, P., V. Berthelot, E. Pottier and J. Normand (2007). Effet of linseed on fatty acid composition of muscles and adipose tissues of lambs with emphasis on trans fatty acids. J. Meat Sci., 77: 678–688.
- Benchaar, C., G.A. Romero-Pérez, P.Y. Chouinard, F. Hassanat, M. Eugene, H.V. Petit and C. Côrtes, (2012). Supplementation of increasing amounts of linseed oilto dairy cows fed total mixed rations: Effects on digestion, ruminal fermentation characteristics,protozoal populations, and milk fatty acid composition. J. Dairy Sci., 95: 4578–4590.
- Broudiscou, L., S. Pochet and C. Poncet (1994). Effect of linseed oil supplementation on feed degradation and microbial synthesis in the rumen of ciliate-free and refaunated sheep. J. Anim. Feed Sci. Technol., 49: 189–202.
- Castañeda-Gutiérrez, E., M.J. De Veth, A.L. Lock, D.A. Dwyer, K.D. Murphy and D.E. Bauman (2007). Effect of supplementation with calcium salts of fish oil on n-3 fatty acids in milk fat. J. Dairy Sci., 90: 4149–4156.
- Chichlowski, M.W., J.W. Schroeder, C.S. Park, W. L. Keller, and D.E. Schimek (2005). Altering the fatty acids in milk fat by including canola seed in dairy cattle diets. J. Dairy Sci., 88: 3084–3094.
- Chikunya, S., G. Demirel, M. Enser, J. D. Wood, R. G. Wilkinson and L. A. Sinclair (2004). Biohydrogenation of dietary n-3 PUFA and stability of ingested vitamin E in the rumen, and their effects on microbial activity in sheep. British J. Nutrition., 91: 539–550.
- Chilliard, Y., C. Martin, J. Rouel and M. Doreau (2009). Milk fatty acids in dairy cows fed whole crude linseed, extruded linseed, or linseed oil, and their relationship with methane output. J. Dairy Science., 92: 5199–5211.
- Chouinard, P.Y., V. Girard and G.J. Brisson (1997). Performance and profiles of milk fatty acids of cows fed full fat, heat-treated soybeans using various processing methods. J. Dairy Sci., 80: 334-342.
- Corl, B. A., L. H. Baumgard, D. A. Dwyer, J. M. Griinari, B. S. Phillips and D. E. Bauman (2001). The role of (9-desaturase in the production of *cis*-9, *trans*-11 CLA. J. Nutr. Biochem., 12:622–630.
- Crawford, R.J. Jr. and W.H. Hoover (1984). Effects of particle size and formaldehyde treatment of soybean meal on milk production and composition for dairy cows. J. Dairy Sci., 67: 1945–1952.
- Deaville, E.R., D.I. Givens and J.S. Blake (2004). Dietary supplements of whole linseed and vitamin E to increase levels of α-linolenic acid and vitamin E in bovine milk. J. Anim. Res., 53: 3–12.
- Doreau, M., D. Bauchart and Y. Chilliard (2011). Enhancing fatty acid composition of milk and meat through animal feeding. J. Animal Production Science., 51: 19–29.
- Doreau, M. and A. Ferlay (2015). Linseed: a valuable feedstuff for ruminants. OCL 2015, 22(6) D611
- El-Essawy, A. M (2019). Effect of lipid source; Linseed or soybean in diets, on rumen and blood fatty acids profiles in Damascus goats. Australian J. Basic and Applied Sciences., 13(5): 1-10.
- Ferraz, T.P.L., M.C. Fiuza, M.L.A. dos Santos, L. Pontes de Carvalho and N.M. Soares (2004). Comparison of six methods for the extraction of lipids from serum in terms of effectiveness and protein preservation. J. Biochem. Biophys. 58: 187–193.
- Fievez, V., B. Vlaeminck, T. Jenkins, F. Enjalbert and M. Doreau (2007). Assessing rumen biohydrogenation and its manipulation in vivo, in vitro and in situ.Eur. J. Lipid Sci. Technol., 109:740–756.
- Fiorentini, G., I.P.C. Carvalho, J.D. Messana, R.C. Canesin, P.S. Castagnino, J.F. Lage, P.B. Arcuri1 and T.T. Berchielli (2015). Effect of Lipid Sources with Different Fatty Acid Profiles on Intake, Nutrient Digestion and Ruminal Fermentation of Feedlot Nellore Steers. Asian Australas. J. Anim. Sci., 28 (11): 1583-1591.

- Focant, M., E. mignolet, M. marique, F. clabots, T. breyne, D. dalemans and Y. larondelle (1998). The effect of vitamin E supplementation of cow diets containing rapeseed and linseed on the prevention of milk fat oxidation. J. dairy sci., 81:1095–1101.
- Gawad, R.M.A., M. Strabel, S.A. Abo El-Nor, H.M. Kattab, A. Cieślak, S.M. Kholif and M. Elnashar (2015). Encapsulation Method to Protect Unsaturated Fatty Acids from Rumen Biohydrogenation In Vitro. J. JIPBS., 2 (3): 240-251.
- Gomez-Cortes, P., C. Tyburczy, J.T. Brenna, M. Jua´ rez and M.A. de la Fuente (2009). Characterization of cis-9, trans-11, trans-15-C18:3 in milk fat by GC and covalent adduct chemical ionization tandem MS. J. Lipid Research., 50:2412–2420.
- Gonthier, C., A.F. Mustafa, D.R. Ouellet, P.Y. Chouinard and H.V. Petit (2005). Feeding micronized and extruded flaxseed to dairy cows: Effects on blood parameters and milk fatty acid composition. J. Dairy Sci., 88:748-756.
- Gravert, H.O. (1987). Dairy Catlle Production. Institute for Milk Production. Federal Dairy Research Centre Kiel FRG. Elsevier Sci. Publ. B.V. New York.
- Jenkins, T and F. Lundy (2001). Feeding various fat sources to lactating dairy cows and their effects on milk quality. available at: http://extension.psu.edu/animals/import/dairy/nutrition/pdf/jenkinsfatsources-and-effects on-milk quality [Accessed Mar 10, 2015].
- Kennelly, J. J. (1996). The fatty acid composition of milk fat as influenced by feeding oilseeds. J. Anim. Feed Sci. Technol., 60:137–152.
- Kholif, A.E., H.M. Khattab, A.A. El-Shewy, A.Z.M. Salem, A.M. Kholif. M.M. El-Sayed, H.M. Gado and M.D. Mariezcurrena (2014). Nutrient digestibility, ruminal fermentation activities, serum parameters and milk production and composition of lactating goats fed diets containing Rice straw treated with pleurotus ostreatus. Asian Australasian J. of Animal Science. 27(3): 357-364.
- Kholif, S. M., T.A. Morsy, O.H. Matloup, H.M. Ebeid and A. M. Kholif (2015). Effects of Crushed Linseed or Linseed Oil Supplementation on Performance of Dairy Goats and Fatty Acid Profile in Milk. J. Life Science., 12(2s).
- Kim, S. C., A. T. Adesogan, L. Badinga and C. R. Staples (2007). Effects of dietary n-6/n-3 fatty acid ration on feed intake, digestibility and fatty acid profiles of the ruminal contents, liver, and muscle of growing lambs. J Anim Sci., 85: 706–716.
- Kim, Y.K., D.J. Schingoethe, D.P. Casper and F.C. Ludens (1993). Supplemental Dietary Fat from Extruded Soybeans and Calcium Soaps of Fatty Acids for Lactating Dairy Cows. J. Dairy Sci., 76: 197-204.
- Knapp, D.M., R.R. Grummer and M.R. Dentine (1991). The response of lactating dairy cows to increasing levels of whole roasted soybeans. J. Dairy Sci., 74: 2563-2572.
- Konitzer, K. and S. Voigt (1963). Direct determination of ammonium in blood and tissue extracts by means of the phenol by chlorite reaction. Clin. Chim. Acta,8:5-11
- Liu, Z.L., D.P. Yang, P. Chen, S.B. Lin, X.Y. Jiang, W.S. Zhao, J.M. Li and W.X. Dong (2008). Effect of dietary sources of roasted oilseeds on blood parameters and milk fatty acid composition. Czech J. Anim. Sci., 53 (5): 219–226.
- Lundy, F. P., E. Block, W. C. Bridges, J. A. Bertrand and T. C. Jenkins (2004). Ruminal biohydrogenation in Holstein cows fed soybean fatty acids as Amides or calcium salts. J. Dairy Sci., 87:1038-1046.
- Miller, W.F., J.E. Shirley, E.C. Titgemeyer and M.J. Brouk (2009). Comparison of full-fat corn germ, whole cottonseed, and tallow as fat sources for lactating dairy cattle. J. Dairy Sci., 92: 3386-3391.
- Nogueira, R.G.S., F.P. Junior, A.S.C. Pereira and P. H. M. Rodrigues (2019). Nutrient digestibility and changes in feeding behavior of cattle fed cottonseed and vitamin E. J. Sci. Agric., 76(2): 112-122.
- NRC (2007). Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids and New World Camelids. The National Academic Press, Washington, DC, USA.
- Petit, H. V (2002). Digestion, milk production, milk composition, and blood composition of dairy cows fed whole flaxseed. J. Dairy Sci., 85:1482–1490.

### Egyptian J. Nutrition and Feeds (2019)

- Petit, H. V (2003). Digestion, milk production, milk composition, and blood composition of dairy cows fed formaldehyde treated flaxseed or sunflower seed. J. Dairy Sci., 86: 2637–2646.
- Petit, H.V and N. Gagnon (2009). Milk concentrations of the mammalian lignans enterolactone and enterodiol, milk production, and whole tract digestibility of dairy cows fed diets containing different concentrations of flaxseed meal. J. Anim. Feed Sci. Technol., 152: 103-111.
- Petit, H.V., M. Ivan and P.S. Mir (2005). Effects of flaxseed on protein requirements and N excretion of dairy cows fed diets with two protein concentrations. J Dairy Sci., 89: 1755–1764.
- Petit, H. V., G. F. Tremblay, E. Tremblay and P. Nadeau (2002). Ruminal biohydrogenation of fatty acids, protein degradability, and dry matter digestibility of flaxseed treated with different sugar and heat combinations. Can. J. Anim. Sci., 82:241–250.
- Pires, A.V., M.L. Eastridge and J.L. Firkins (1996). Roasted soybeans, blood meal, and tallow assources of fat and ruminally undegradable proteinin the diets of lactating cows. J Dairy Sci., 79: 1603-1610.
- Price, P., C. Nelson, S.P. Clark (2000). Omega–3 polyunsaturated fatty acid regulation of gene expression. J. Curr Opin Lipidol.,11:3–7.
- Radivojević, M., G., H. Grubić, Šamanc, M. Adamović and N. Đorđević (2011). Heat treated soybeansin the nutrition of high producing dairy cows. Afr. J. Biotechnol., 10 (19): 3929-3937.
- Rice, B.H., J. Kraft, F. Destaillats, D.E. Bauman and A.L. Lock (2012). Ruminant-produced trans-fatty acids raise plasma HDL particle concentrations in intact and ovariectomized female hartley guinea pigs. J. Nutr., 142: 1142–1151.
- Ruxton, C (2007). The health benefits of omega-3polyunsaturated fatty acids: a review of the evidence. Commentary. J. Hum. Nutr. Diet., 20: 275–287.
- Sangeetha, R., I. Arulpandi and A. Geetha (2011). Bacterial lipases as potential industrial biocatalysts: An overview. J. Of Microbiology., 6(1): 1-24.
- Santos, N.W., G.T. Santos, D.C. Silva-Kazama, P.A. Grande, P.M. Pintro, F.E. de Marchi, C.C. Jobim and H.V. Petit (2014). Production, composition and antioxidants in milk of dairy cows fed diets containing soybean oil and grape residue silage. J. Liv. Sci., 159: 37–45.
- SAS (2006). SAS Institute.User's Guide: Statistics. Ver 9.0. SAS Institute, Cary, N.C., USA.
- Scollan, N. D., M. S. Dhanoa, N. J. Choi, W. J. Maeng, M. Enser and J. D. Wood (2001). Biohydrogenation and digestion of long chain fatty acids in steers fed on different sources of lipids. J. Agric. Sci. (Camb.) 136:345–355.
- Scott, T.W., L.J. Cook and S.C. Mills (1971). Protection of dietary polyunsaturated fatty acids against microbial hydrogenation in ruminants. J. Amer Oil Chem Soc., 48:358–364.
- Sharma, H.R., J.R. Ingalls and J. A. Mckirdy (1972). Nutritive value of formaldehyde-treated rapeseed meal for dairy calves. Can. J. Anim. Sci.52:363 –371.
- Shingfield, K.J., S. Ahvenjarvi, V.A. Toivonen, A. Arola, K.V.V. "Nurmela, P. Huhtanen and J.M. Griinari (2003). Effect of dietary fish oil on biohydrogenation of fatty acids and milk fatty acid content in cows. J. Animal Science., 77: 165–179.
- Suksombat, W., L.P.Thanh, C. Meeprom and R. Mirattanaphrai (2014). Effects of Linseed Oil or Whole Linseed Supplementation on Performance and Milk Fatty Acid Composition of Lactating Dairy Cows. Asian Australas. J. Anim. Sci., 27:951-959.
- Tymchuk, S.M., G.R. Khorasani and J.J. Kennelly (1998). Effect of feeding formaldehyde- and heattreated oil seed on milk yield and milk composition. Can. J. Anim. Sci., 78:693-700.
- Van Soest, P.J., G.B. Robertson and B.A. Lewis (1991). Symposium: Carbohydrate methodology, metabolism, and nutritional implications in dairy cattle. J. Dairy Science., 74: 3583–3597.
- Wachira, A. M., L. A. Sinclair, R. G. Wilkinson, K. Hallett, M. Enser and J. D. Wood (2000). Rumen biohydrogenation of n-3 polyunsaturated fatty acids and their effects on microbial efficiency and nutrient digestibility in sheep. J. Agric. Sci., 135:419–428.
- Warner, A. C. J. (1964). Production of volatile fatty acids in the rumens methods of measurements. J. Nut. Abst. Rev., 34: 39.

- Weill, P., B. Schmitt, G. Chesneau, N. Daniel, F. Safraou, and P. Legrand (2002). Effects of Introducing Linseed in Livestock Diet on Blood Fatty Acid Composition of Consumers of Animal Products. J. Ann Nutr Metab., 46:182–191.
- Zened, A., A. Troegeler-Meynadier, T. Najar and F. Enjalbert (2012). Effects of oil and natural or synthetic vitamin E on ruminal and milk fatty acid profiles in cows receiving a high-starch diet. J. Dairy Sci., 95 :5916–5926.

تأثير إضافة كبسولات زيت بذرة الكتان مع فيتامين هـ علي أداء و مشتقات الدم و كمية اللبن في الماعز الحلابة

عبير محد عبد الحليم العيسوي1 ، إبراهيم محد عبد الحافظ خطاب2 ، أحلام رمضان عبده 1 و عادل محد عبد الواحد1

<sup>1</sup> قسم تغذية الحيوان- شعبة الإنتاج الحيواني و الدواجن- مركز بحوث الصحراء- المطرية- القاهرة- مصر.
<sup>2</sup> قسم الإنتاج الحيواني و السمكي- كلية الزراعة الصحراوية و البينية- جامعة مطروح- مطروح- مصر.

تهدف هذه الدراسة ألى تقييم تأثير إضافة كبسولات زيت بذرة الكتان فقط أو مع فيتامين هـ كإضافات غذائية علي أداء الحيوان و تركيب الأحماض الدهنية في بلازما الدم و في اللبن أثناء فترة الرضاعة. تم تقسيم 24 من المعز الدمشقي العشار في منتصف الحمل (متوسط وزن 44.5 ± 1.2 كجم) الي ثلاث مجموعات (8 ماعز للمجموعة). المجموعة الأولي (المقارنة) تغذت علي دريس البرسيم و مخلوط العلف المركز بنسبة 50:50 بدون اي إضافات ، المجموعة الثانية تغذت علي نفس غذاء مجموعة المقارنة + 2.5 جم كبسو لات زيت بذرة الكتان لكل حيوان و المجموعة الثالثة تغذت مثل مجموعة المقارنة + 2.5 جم كبسولات زيت الكتان + 600 وحدة دولية من فيتامين هـ لكل حيوان. استمرت التجربة 135 يوم من منتصف الحمل حتى 60 يوم بعد الولادة . أظهرت النتائج أن إضافة كبسولات زيت الكتان نتج عنها زيادة معنوية كبيرة في كمية اللبن و كمية الدهون و البروتين و سُكر اللاكتوز و المكونات الصلبة باللَّبن و كذلك النسبة المئوية للدهن مقارَّنة بمجموعة المقارنة. ارتفَّع مستوي حمض الأوميجا ۔3 ببلازما الدم مع الكبسولات و كذلك أحد احماض الأوميجا ۔6 مع إضافة الفيتامين. و في اللبن :إنخفض مستوي الأحماض الدهنية المشبعة بينما ارتفع مستوى الأحماض غير المشبعة الأحادية و المتعددة التشبع مع إضافة الكبسولات مما نتج عنه انخفاض النسبة الكلية للأحماض الدهنية المشبعة و أحماض الأوميجا-6 الكلية و النسبة بين أوميجا-6 : أوميجا-3 مع ارتفاع نسبة الأحماض الدهنية الغير مشبعة و أحماض الأوميجا -3 . مما ترتب عليه إنتاج لبن صحي و أكثر فائدة بدرجة كبيرة. أيضا تحسنت بعض مشتقات الدم حيث انخفض مستوى الكوليستيرول و الكرياتينين و مستوى انزيم الليبيز مقارنة بمجموعة المقارنة. و علي الجانب الأخر فقد ارتفعت في الدم نسبة البولينا و الدهون الثلاثية والبروتينات الدهنية قليلة و عالية الكثافة و مضادات الأكسدة الكلية بينما انخفض نشاط انزيم الليبيز مع إضافة فيتامين هـ. نستخلص مما سبق ان إضافة كبسولات زيت الكتان أسهمت في زيادة انتاج اللبن نو مواصفات صحية مرتفعة للمستهلك لأن زيت الكتان يرفع من القيمة الغذائية للبن بزيادة الأحماض الدهنية المفيدة به. تبين ان إضافة فيتامين ه مع الكبسولات أدي الي تحسن في منتجات الماعز محل الدر اسة. لهذا فمن المهم زيادة الأبحاث و الدر اسات على الحيوان للوصول للمستوي المناسب لمنتج أفضل من كبسو لات زيّت الكتان و الفيتامين.